



Journal of Plant Nutrition

ISSN: 0190-4167 (Print) 1532-4087 (Online) Journal homepage: <http://www.tandfonline.com/loi/lpla20>

Effect of nutrient solution salinity and ionic concentration on parsley (*Petroselinum crispum* Mill.) essential oil yield and content

Juan Eugenio Álvaro, María Teresa Lao, Miguel Urrestarazu, Mourad Baghour & Moumen Abdelmajid


To cite this article: Juan Eugenio Álvaro, María Teresa Lao, Miguel Urrestarazu, Mourad Baghour & Moumen Abdelmajid (2016) Effect of nutrient solution salinity and ionic concentration on parsley (*Petroselinum crispum* Mill.) essential oil yield and content, Journal of Plant Nutrition, 39:8, 1057-1062, DOI: [10.1080/01904167.2015.1061552](https://doi.org/10.1080/01904167.2015.1061552)

To link to this article: <https://doi.org/10.1080/01904167.2015.1061552>



Accepted author version posted online: 06 Jul 2015.
Published online: 06 Jul 2015.



Submit your article to this journal 



Article views: 191



View Crossmark data 

Effect of nutrient solution salinity and ionic concentration on parsley (*Petroselinum crispum* Mill.) essential oil yield and content

Juan Eugenio Álvaro^a, María Teresa Lao^b, Miguel Urrestarazu^{b,c}, Mourad Baghour^d, and Moumen Abdelmajid^d

^aPontificia Universidad Católica de Valparaíso, Valparaíso, Chile; ^bDepartment of Agronomy, University of Almería, Almería, Spain; ^cUniversidad de Tarapacá, Arica, Chile; ^dFaculté Pluridisciplinaire de Nador, Université Mohamed Premier, Nador, Morocco

ABSTRACT

The growth and essential oil (EO) production of parsley were evaluated in response to salinity and nutrient solution concentrations in a soilless culture. Parsley plants that were 60 days old were potted in a coconut fiber and peat moss medium and were treated with four different nutrient solutions, including T1, T2, T3 and T4. The T1 nutrient solution was the standard, the T2 and T3 solutions contained incremental macronutrient concentrations with an electrical conductivity (EC) of up to 2.2 and 3.2 dS m⁻¹, respectively, and the T4 solution was the same as T2 but with sodium chloride (NaCl) and an incremental macronutrient concentration with an EC of 3.2 dS m⁻¹. Next, these plants were grown for 90 days in a greenhouse with natural daylight in Nador, Morocco. Shoot and root growth significant decreased with increasing EC. However, the salinity that resulted from the addition of NaCl did not affect plant growth in the nutrient solutions. The optimum obtained growth and EO production were 1.2 and 2.2 dS m⁻¹, respectively. Consequently, the optimum EC value (based on the EO production) of parsley in the soilless culture was 1.2–2.2 dS m⁻¹.

ARTICLE HISTORY

Received 21 September 2013
Accepted 25 November 2014


KEYWORD

Salinity; saline stress; medicinal and aromatic plants; growth and development; salt tolerance; soilless culture; coconut fiber; specific salinity effect

Introduction

Parsley, *Petroselinum crispum* Mill. (Umbelliferae), is a biennial plant that is widely used to garnish and aromatize foods (as fresh green sprigs) and as a medicinal and aromatic plant. Parsley is grown throughout most of the world.

Since the middle ages, essential oils have been widely used for bactericidal, virucidal, fungicidal, antiparasitic, insecticidal, medicinal and cosmetic applications, especially in the pharmaceutical, sanitary, cosmetic, agricultural, and food industries (Bakkali et al., 2008). Regarding crop protection, plant essential oils have been recently used against specific pests and fungi (Daferera et al., 2003; Isman, 2003). Parsley (*Petroselinum crispum* Mill) contains flavonoids (apiin and luteolin) and essential oils (apiol and myristicin) that are responsible for the medical uses and toxicity of parsley. Furanocoumarins (psoralen, bergapten, isoimperatorin, oxypeucedanin, xanthoxin, trioxsalen and angelicin) are also an important chemical component of parsley plants (Mimica-Dukiae and Popoviae, 2007). Furthermore, myristicin (63.9%) and apiol (14.4%) are major constituents of parsley. However, 34 additional essential oil compounds have been found in parsley (Pino et al., 1997).

CONTACT Miguel Urrestarazu ✉ mgavilan@ual.es  Department of Agronomy, University of Almería, La Cañada de San Urbano s/n, C.P. 04120, Almería, Spain.

Color versions of one or more of the figures in the article can be found online at www.tandfonline.com/lpla.

© 2016 Taylor & Francis Group, LLC

Since the 1960s, the influence of salinity on horticulture (Sonneveld and Voogt, 2009) and aromatic plants (Sonneveld et al., 1999) has been studied.

The scarce availability of water in the Mediterranean basin is one of the main factors that will limit agricultural development, particularly between 2000 and 2025 (Chartzoulakis et al., 2002). To overcome water shortages and to satisfy the increasing water demands of agricultural development, the use of marginal quality water (brackish, reclaimed, and drainage) will become necessary in many countries (Correia, 1999; Hamdi et al., 1995). However, the use of saline water for irrigation requires an adequate understanding of how salts affect plant performance.

The increasing demand for irrigation water is a worldwide problem. Additionally, poor quality water is a resource that has not been fully exploited, especially in semi-arid zones. In semi-arid zones where water quality is poor, the efficiency of nutrient use must be enhanced by adjusting the ion concentrations of the nutrient solutions. This task must be performed without decreasing production and by avoiding contaminant emissions as much as possible (Urrestarazu et al., 2008).

Many commercial greenhouse industries are forced to use poor quality water due to the presence of sodium chloride (NaCl). Occasionally, low to moderate levels of salinity are achieved in soilless culture by adding NaCl or major nutrients to improve fruit quality (Adams, 1991). This cultural management method could potentially be applied to essential oil production.

Contrasting reports are present in the literature regarding the response of essential oils to salt stress (15). For example, low and moderate saline stress has been shown to affect fennel (Abd El-Wahab, 2006) and marjoram (Baatour et al., 2010) negatively. In contrast, other authors recorded lower EO production when aromatic and medicinal plants were subjected to stress. For example, Baher et al. (2002) reported that the essential oil (EO) production of *Satureja hortensis* plants decreased when they were subjected to water stress. Similar results were obtained for *Lipia citriodora* under salt stress (Tabatabaie and Nazari, 2007). In contrast, the same authors reported that EO production in *Menhta piperita* did not decrease under the same saline conditions.

Therefore, the beneficial or detrimental effects of salt stress or nutrient solutions that are enriched with NaCl are different dependent on the type of plant tested.

In contrast, the advantages of soilless culture for growing medicinal plants suggest that these cultivation systems could be a powerful tool for the medicinal product industry (Hyden, 2006).

Very few studies have focused on the effects of nutrient solutions on soilless aromatic or medicinal plant crop from the following perspectives: 1) growth [e.g., Urrestarazu et al. (2013) using thyme lime], 2) biomass and essential oil production [e.g., Maia et al., (1999) using garden mint; Cordovilla et al. (2013) using thyme and lavender] or 3) essential oil production (based on biomass) and the ratio or concentration of each essential oil in peppermint and lemon verbena plants] (Tabatabaie and Nazari, 2007) (Table 1).

The objective of this experiment was to determine the effects of a nutrient solution's electrical conductivity (EC) and salinity (NaCl) on the growth and essential oil production of parsley.

Table 1. Examples of nutrient solution manipulation experiments in aromatic and medicinal crops grown in soilless culture.

Plant	Reference	Biomass yield	Essential oil (EO) production	Component (EO)	Remark
<i>Lipia citriodora</i> var. Verbena	Tabatabaie and Nazari (2007)	Yes	Yes	Yes	Level of EC and NaCl
<i>Mentha crispera</i>	Maia et al. (1999)	Yes	Si	No	Fertigation Method
<i>Mentha piperita</i> var. officinalis	Tabatabaie and Nazari (2007)	Yes	Yes	Yes	Level of EC and NaCl
<i>Petroselinum crispum</i>	Chondraki et al. (2012)	Yes	No	No	Level of EC by NaCl under NFT
<i>Satureja hortensis</i>	Najafi et al. (2010)	Yes	Yes	Yes	Level of EC and NaCl. Water culture
<i>Thymus citriodorus</i>	Urrestarazu et al. (2013)	Yes	No	No	Level of EC and NaCl
<i>Thymus vulgaris</i>	Cordovilla et al. (2013)	Yes	Yes	No	Level of EC and NaCl
<i>Lavandula angustifolia</i>	Cordovilla et al. (2013)	Yes	Yes	No	Level of EC and NaCl

Materials and methods

A 60-day-old parsley (*Petroselinum crispum* Mill.) plant (from seed) was potted in a 1 L pot. Each pot was filled with a mixture of coconut fiber and commercial peat moss (2:1; vol:vol). The experiment was conducted in a plastic greenhouse at Mohamed Premier University (Nador, Morocco) for 90 days. Four different treatments were established on 1 April. The control treatment (T1) consisted of a standard nutrient solution with an EC of 1.2 dS m^{-1} , which was similar to the nutrient solution described by Sonneveld and Straver (1994) for pot cultivation. Treatments T2 and T3 had incremental micronutrient concentrations with an electrical conductivity (EC) of up to 2.2 and 3.2 dS m^{-1} , respectively, using a concentration nutrient solution on base of macronutrients. Treatment T4 was the same as treatment T2, but was adjusted with NaCl to have an EC of up to 2.3 dS m^{-1} .

Ninety days after transplanting, the plants were gently removed from their substrate. Next, the roots were washed with distilled water and the plant roots and shoots were divided for each treatment and replicate. Each sample was placed in a forced-air oven at 85°C until a constant weight was achieved. The weight was recorded with a scale that had a precision of 0.01 g.

The essential oil (EO) concentration was determined by hydro distillation of the aerial part (shoot) of the air-dried plants (500 g) with a modified Clevenger apparatus after 4 hours. These measurements began when the first drop of liquid condensed in the cooling column and dripped into the grade burette. EO production was calculated by adding the volume/weight ratio (vol/w) of the cold oil collected that was collected from the burette to the original material.

The experiment was completely randomized with 16 replicate and treatment pots (8 pots for growth and 8 pots for EO). A Tukey's multiple range test at $P \leq 0.05$ was used to differentiate the means. The experimental design and data analysis were based on the procedure described by Little and Hills (1987). The Statgraphics® Plus 5.0 statistical package was used to process the data (Statistical Graphics Corp., Rockville, MD, USA).

Results and discussion

The saline treatments negatively affected vegetative growth (at $P \leq 0.05$, Figure 1). Specifically, the growth was reduced by 30 and 40% for the dry and fresh weights of the shoots, respectively. Even for relatively moderate EC values (2.2 dS m^{-1}), which could be considered to be optimal for some aromatic plants such as Freisia (Sonneveld and Straver, 1994), plant production was reduced. This significant biomass reduction is important because a large portion of parsley plant production is marketed worldwide for seasoning fresh meals. These data are consistent with previously observed peppermint and lemon verbena biomass reductions with varying EC from 1.4 to 2.8 dS m^{-1} (Tabatabaie and Nazari, 2007) and with the results obtained by Urrestarazu et al. (2013) for lime thyme leaves. The effects of specific salt toxicity on the roots were not clear. The saline treatments with macronutrient concentrations (T2 and T3) had lower root growth (up to 50%); however, treatment T4 had the same root biomass as treatment T2. Because the roots are not commercialized and contain only small amounts of EO (approximately 0.1%) (Tyler et al., 1988), their biomass is less important from a commercial standpoint. The response of the parsley to the treatment suggested that it is highly sensitive to salinity (Figure 2).

When comparing the biomass weight of the plants grown in the nutrient solution with those grown in the concentrated macronutrient solution (T3) and NaCl (T4), a reduction in root and shoot growth was observed. Tabatabaid and Nazari (2007) observed the same effect for peppermint plants, and Urrestarazu et al. (2013) observed the same effect for lime thyme plants. Tabatabaid and Nazari (2007) observed that NaCl affected lemon verbena by significantly reducing its biomass when the EC increased due to the addition of NaCl but not macronutrients. This specific saline effect due to NaCl is well described by Sonneveld (2004) and Sonneveld and Voogt (2009).

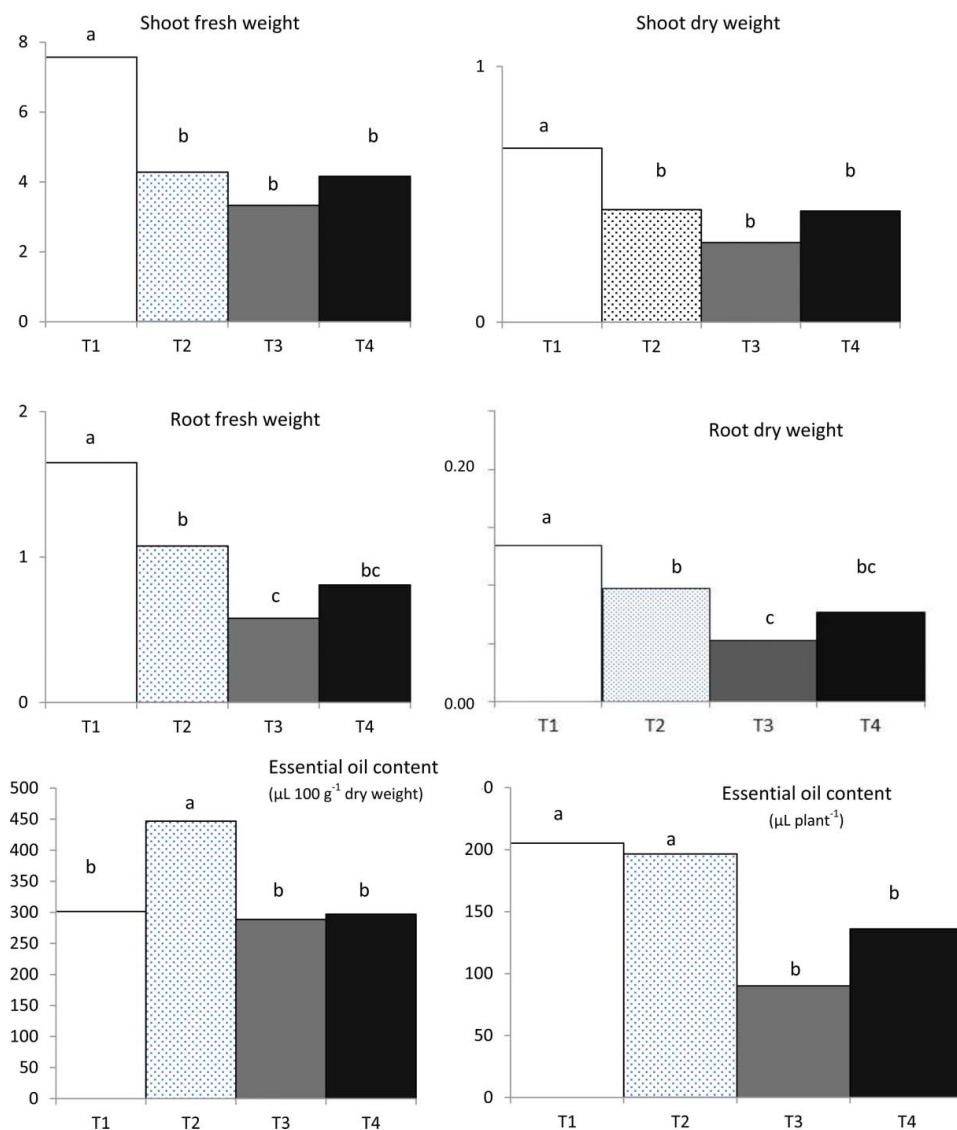


Figure 1. Growth parameter (g plant⁻¹) and essential oil contents of parsley. T1, T2, T3 and T4 indicate nutrient solution treatments with electric conductivities of 1.2, 2.2, 3.2 from concentrated nutrient solution on base of macronutrients, and 3.2 (with 6 mM NaCl), respectively. Different letters indicate significant differences ($P \leq 0.05$).

The EO concentrations that were obtained for the parsley shoot were similar to those reported by Tyler et al. (1998) and Orav et al. (2003).

The highest oil concentrations were obtained at moderate salinity values ($EC = 2.2 \text{ dS m}^{-1}$, T2). No significant differences between the low (1.2 dS m^{-1}) and high (3.2 dS m^{-1} , T3 and T4) conductivity treatments were observed. However, because the greatest biomass was registered for treatment T1, it was concluded that the optimal EO production by the plants occurs at an EC of between 1.2 and 2.2 dS m^{-1} . Similar results were previously reported for peppermint, lemon verbena (Tabatabaie and Nazari, 2007) and lavender (Maia et al., 1999). Therefore, under very controlled conditions (as in soilless cultures) and when the tolerance and response of a plant to specific salinity levels are known, fertigation (composition of the nutrient solution) can be controlled. Depending on their aim, produces can 1) improve biomass development, 2) improve EO productivity or 3) optimize these first two factors.

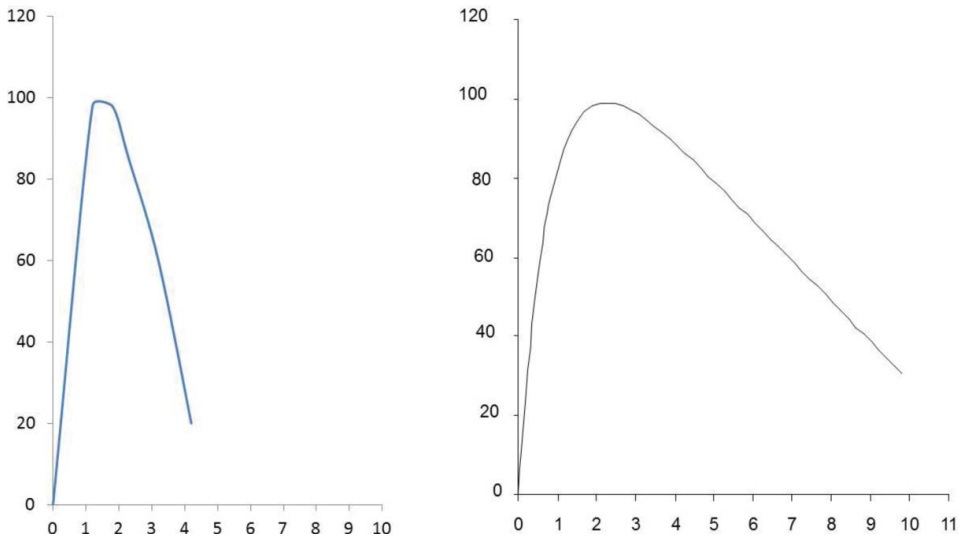


Figure 2. Comparative mean yield (%) of parsley and chrysanthemum with the nutrient solution EC (dS m^{-1}) [Sonneveld (2004) and Sonneveld et al. (2005)].

Conclusions

The results suggest that an EC of 1.2 dS m^{-1} is optimal for obtaining high economic yields, particularly for biomass yields (which have basic several uses). However, moderate EC values between 1.2 and 2.2 are beneficial for parsley crop essential oil production in soilless cultivation.

Funding

This work was supported by the Spanish Ministry of Education and Science Project FEDER AGL2015-67528-R) and the author gratefully acknowledge the support of CONICYT through Project FONDECYT DE INICIACIÓN no. 11140154.

References

- Abd El-Wahab, M. A. 2006. The efficiency of using saline and fresh water irrigation as alternating methods of irrigation on the productivity of *Foeniculum vulgare* mill subsp. Vulgare var. Vulgare under north sinai conditions. *Research Journal of Agricultural and Biological Sciences* 2: 571–577.
- Adams, P. 1991. Effect of increasing salinity of the nutrient solution with major nutrient or sodium chloride on the yield, quality and composition of tomato grown in rockwool. *Journal of Horticultural Science and Biotechnology* 66: 201–207.
- Baatour, O. R., W. Kaddour, A. Wannes, M. Lachaal, and B. Marzouk B 2010. Salt effects on the growth, mineral nutrition, essential oil yield and composition of marjoram (*Origanum majorana*). *Acta Physiologiae Plantarum* 32: 45–51.
- Baher, Z. F., M. Mirza, M. Ghorbanli, and M. Bagher Rezaii. 2002. The influence of water stress on plant height, herbal and essential oil yield and composition in *Satureja hortensis*. *Flavour and Fragrance Journal* 17: 275–277.
- Bakkali, F., S. Averbeck, D. Averbeck, and M. Idaomar. 2008. Biological effects of essential oils – a review. *Food and Chemical Toxicology* 46: 446–475.
- Chartzoulakis K., M. Loupassaki, M. Bertaki, and M. Androulakis. 2002. Effects of NaCl salinity on growth, ion content and CO₂ assimilation rates of six olive cultivars. *Scientia Horticulturae* 96: 235–247.
- Cordovilla, P., M. Bueno, C. Aparicio, and M. Urrestarazu. 2014. Effects of salinity and the interaction between *Thymus vulgaris* and *Lavandula angustifolia* on growth, ethylene production and essential oil contents. *Journal of Plant Nutrition* 37: 875–888.
- Correia, F. N. 1999. Water resources in the Mediterranean region. *Water International* 24: 22–30.
- Daferera, D. J., B. N. Ziogas, and M. G. Polissiou. 2003. The effectiveness of plant essential oils on the growth of *Botrytis cinerea*, *fusarium* sp. and *Clavibacter michiganensis* subsp *michiganensis*. *Crop Protection* 22: 39–44.
- Hamdi, A., M. F. Abud-Zeid, and C. Lacirignola. 1995. Water crisis in the Mediterranean. Agricultural water demand management. *Water International* 20: 176–187.

- Hyden, A. 2006. Aeroponic and hydroponic systems for medicinal herb, rhizome, and root crops. *HortScience* 41: 536–538.
- Isman, B. N. 2000. Plant essential oils for pest and disease management. *Crop Protection* 19: 603–608.
- Little, T. M., and F. G. Hills. 1987. *Statistical Methods for Agricultural Research* Mexico City: Ed. Trillas.
- Maia, N. B., Q. A. Carmello, and M. Marques. 1999. Automatic nutrient solution delivery for hydroponic plant cultivation in pots. *Scientia Agriculturae* 56: 103–110.
- Mimica-Dukić, N., and M. Popović. 2007. *Apiaceae* species. A promising sources of pharmacologically active compounds and *Petroselinum crispum*, *Apium greveolens* and *Pastinaca sativa*. In: *Recent Progress in Medicinal Plant Species*, eds. J. N. Govil, and V. K. Singh Edits, pp. 132–133. Houston, TX: Phytopharmacology and Therapeutic Values III, LLC.
- Orav, A., T. Kailas, and A. Jegorova. 2003. Composition of the essential oil of dill, celery, and parsley from Estonia. *Proceedings of Estonian Academy of Science, Chemistry* 52: 147–154.
- Pino, J. A., A. Rosado, and V. Fuentes. 1997. Herb oil of parsley (*Petroselinum crispum* mill.) from Cuba. *Journal of Essential Oil Research* 9: 241–242.
- Sonneveld, C. 2004. La nutrición mineral y salinidad en los cultivos sin suelo. Su manejo [Mineral vegetal nutrition. Management]. In: *Tratado de Cultivo sin Suelo [Handbook of Soilless Culture]*, ed. M. Urrestarazu, pp. 305–367. Madrid: Mundi-Prensa.
- Sonneveld, C., R. Bass, H. M. C. Nijssen, and J. J. De Hoog. 1999. Salt tolerance of flower crops grown in soilless culture. *Journal of Plant Nutrition* 22: 1033–1048.
- Sonneveld, C., and N. Straver. 1994. *Nutrient Solutions for Vegetables and Flower Grow in Water or Substrates*. Naaldwijk, The Netherlands: Proefstation voor tuinbouw onder glas te Naaldwijk.
- Sonneveld, C., A. L. van den Bos, and W. Voogt. 2005. Modeling osmotic salinity effects on yield characteristics of substrate-grown greenhouse crops. *Journal of Plant Nutrition* 27: 1931–1951.
- Sonneveld, C., and W. Voogt. 2009. *Plant Nutrition of Greenhouse Crops*. Dordrecht, the Netherlands: Springer.
- Tabatabaid, S. J., and J. Nazari. 2007. Influence of nutrient concentrations and NaCl salinity on the growth, photosynthesis, and essential oil content of peppermint and lemon verbena. *Turkish Journal of Agricultural and Forestry* 31: 245–253.
- Tyler, V. E., L. R. Brady, and J. E. Robbers. 1988. *Pharmacognosy*, 9th ed. Philadelphia: Lea & Febiger.
- Urrestarazu, M., L. Borges, S. Burés, and J. E. Álvaro. 2013. Response of lime thyme to salinity and ionic concentration in nutrient solution. *Journal of Plant Nutrition* 36: 562–565.
- Urrestarazu, M., M. C. Salas, D. Valera, and A. Gómez, and P. C. Mazuela. 2008. Effects of heating nutrient solution on water and mineral uptake and early yield of two cucurbits under soilless culture. *Journal of Plant Nutrition* 31: 1532–4087.